AMENDMENTS TO THE CLAIMS

1. (Currently amended) A method for the construction of randomized gene libraries in suitable yeast cells sensitive to *Kluyveromyces lactis* γ toxin and which are capable of homologous recombination comprising the following steps:

introducing into said yeast cells capable of homologous recombination;

- a) a target vector comprising a first DNA sequence coding for at least a γ -subunit of a *Kluyveromyces lactis* killer toxin as negative selection marker, said DNA sequence being flanked at its 5' end by a first target sequence and at its 3' end by a second target sequence and;
- b) a donor DNA sequence which is flanked at its 5' end by a DNA sequence which is homologous to said first target sequence and flanked at its 3' end by a DNA sequence which is homologous to said second target sequence; and

cultivation of said yeast cells under suitable conditions allowing the selection of cells in which said DNA sequence in the target vector encoding at least a γ -subunit of a *Kluyveromyces lactis* killer toxin has been replaced by said donor sequence by means of homologous recombination thereby abolishing expression of said γ -subunit of a *K. lactis* killer toxin.

- 2. (Previously presented) The method of claim 1, wherein said target vector further comprises a second DNA sequence encoding at least one protein region.
- 3. (Original) The method of claim 2 wherein said first DNA sequence of said target vector encoding at least the γ -subunit of the *K. latics* killer toxin and being flanked by said two target sequences replaces a protein region encoding DNA sequence of said second DNA sequence comprised in said target vector.
- 4. (Previously presented) The method of claim 1 wherein said DNA sequence encoding at least the γ subunit of the K. *lactis* killer toxin is under control of a heterologous promoter.

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5. (Original) The method of claim 4 wherein said promoter is located between the DNA sequence encoding at least the γ subunit of *K. lactis* killer toxin and one of the two target sequences.

- 6. (Currently amended) The method of claim 1, wherein said first DNA sequence of said target vector comprises at least one <u>restriction enzyme</u> unique recognition site <u>that is unique</u> for a given restriction enzyme.
- 7. (Previously presented) The method of claim 6, wherein said unique recognition site is located in the coding region of the γ -toxin DNA sequence.
- 8. (Currently amended) The method of claim [[+]] 2 wherein said first second DNA sequence encodes an antibody or a single chain antibody.
- 9. (Previously presented) The method of claim 8 wherein said first DNA sequence of said target vector replaces at least one CDR region of said antibody or said single chain antibody.
- 10. (Original) The method of claims 8 or 9 wherein said first DNA sequence comprising the γ subunit of K. *lactis* killer toxin is transcribed in the opposite direction of said antibody or single chain antibody gene.
- 11. (Previously presented) The method of claim 1 wherein said γ -toxin subunit of the *K. lactis* killer toxin lacks a signal peptide.
 - 12. (Canceled)
- 13. (Previously presented) The method of claim 1 wherein said target vector is introduced into said host cells in linearized form.
- 14. (Currently amended) The method of claim 13 wherein said target vector is linerarized by cutting with a restriction enzyme recognizing in said first DNA sequence of said target vector said at least one unique restriction enzyme recognition site that is unique for a given restriction enzyme.
- 15. (Previously presented) The method of claim 1 wherein said donor sequence comprises a DNA sequence encoding a protein region.

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16. (Previously presented) The method of claim 1 wherein said target vector and said donor sequence are introduced into said host cells by co-transformation.

17. (Previously presented) The method of claim 12 wherein said yeast cells are cultivated at a temperature selected from the range of 24°C to 30°C.

18. (Canceled)

19. (Withdrawn) Use of a *Kluvveromyces lactis* killer toxin γ-subunit as negative selection marker for the construction of randomized gene libraries and/or region replacement by homologous recombination.

20. (Withdrawn) A DNA vector which comprises the following sequences: a first target sequence for homologous recombination, a TEF promoter from Ashbya gossypii driving transcription of a K. lactis killer toxin, a DNA sequence encoding at least a γ-subunit of a K. lactis killer toxin and a second target sequence for homologous recombination.

- 21. (Withdrawn) A host cell comprising a vector of claim 20.
- 22. (Previously presented) The method of claim 4 wherein the promoter is a constitutive promoter.
- 23. (Previously presented) The method of claim 4 wherein the promoter is a TEF promoter from Ashbya gossypii.
- 24. (Currently amended) The method of claim 6 wherein the unique restriction enzyme recognition site that is unique for a given restriction enzyme is located between the coding region of the γ -toxin DNA sequence and the promoter.
- 25. (Previously presented) The method of claim 9 wherein the first DNA sequence of said target vector replaces a CDR3V_L region of said antibody or said single chain antibody.
- 26. (Previously presented) The method of claim 9 where the first DNA sequence of said target vector replaces a CDR2 and a CDR3 region at said antibody or said single chain antibody.

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27. (Previously presented) The method of claim 12 wherein said host cells are *Saccharomyces cerivisiae* cells.

- 28. (Previously presented) The method of claim 15 wherein said donor sequence comprises a DNA sequence encoding a CDR region of an antibody.
 - 29. (Withdrawn) The host cell of claim 20 which is a yeast cell.
 - 30. (Withdrawn) The host cell of claim 29 which is a Saccharomyces cerevisiae.
- 31. (Previously presented) The method of claim 2, wherein said second DNA sequence encodes more than two protein regions.
- 32. (Previously presented) The method of claim 2, wherein said second DNA sequence encodes a full length protein.
- 33. (Previously presented) The method of claim 15, wherein said protein region is a CDR region of an antibody.